



Natural and environmentally friendly way of managing weeds without herbicides

Yoshiharu FUJII, **by Allelopathy**

Tokyo University of Agriculture and Technology,
2.50pm-3.20pm, Monday 24 February 2014

Crown Perth, Burswood



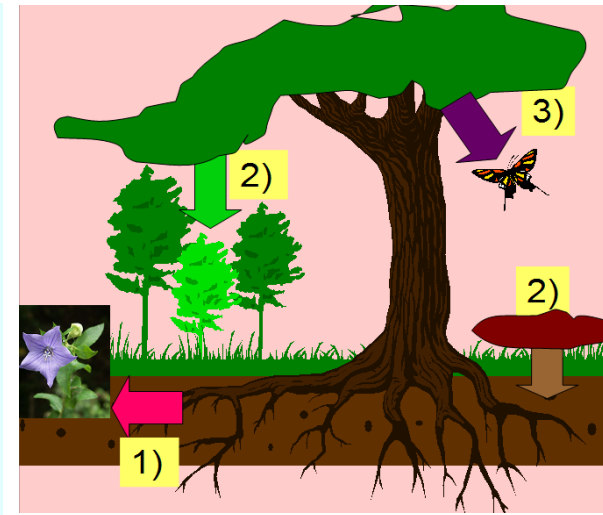
Outline of Talk

What is **Allelopathy**

- Selection of allelopathic plants by **Bioassays**
- New **Concept of Total Activity** for the isolation of Allelochemicals.
- **Cover crop with allelopathic activity** provides natural and environmentally friendly way of managing weeds without herbicides.

Allelopathy

Interaction (inhibitory or stimulatory)
between plants or plant to other life
by natural chemicals
(allelochemicals)



Plant to Plants weed control

Plant to Microorganisms . . . disease control

Plant to Insects harmful Insect control

Plant to Animals nematode, boar, etc.

Allelopathy is the same concepts as “Antibiotics”

Bioassay for Three Route

All these Bioassays are my original Assay

1) Exudation (from root)

→ Plant Box Method

2) Leaching (from leaf or litter)

→ Sandwich Method

3) Volatilization (mainly from leaf as volatile chemicals)

→ Dish Pack Method

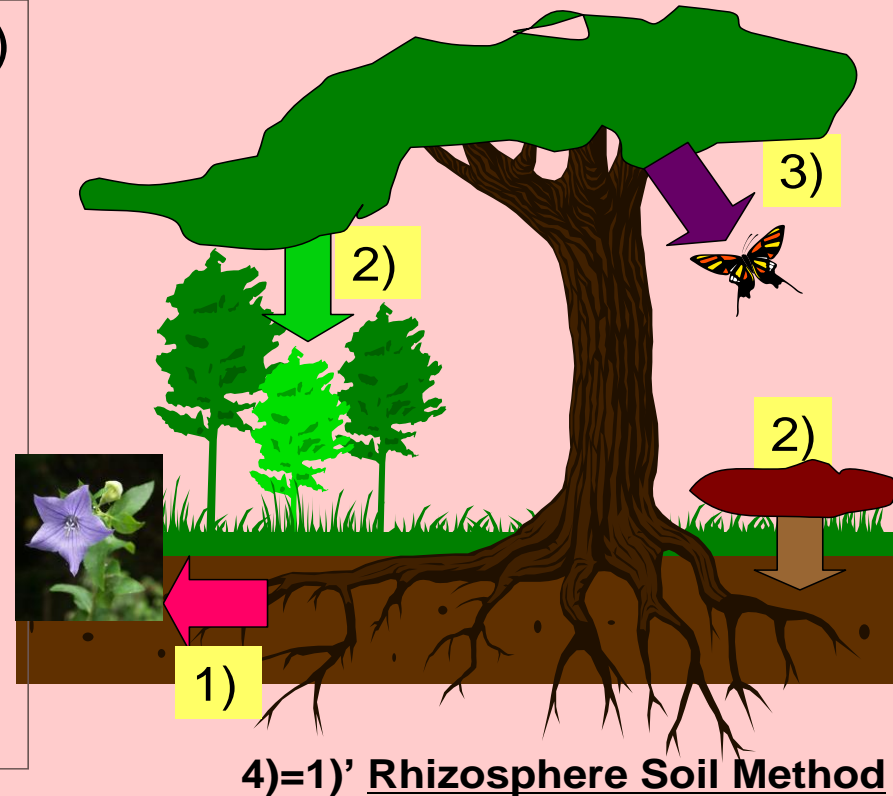


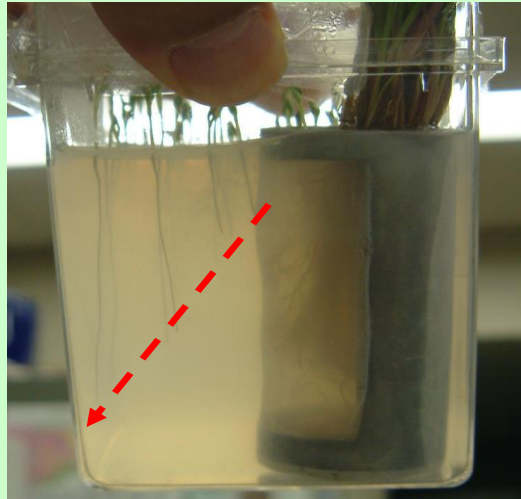
Fig. 1 Route of Allelopathy and Bioassay

1) Specific Bioassays for Allelopathy

For the screening of
Allelopathic Plants

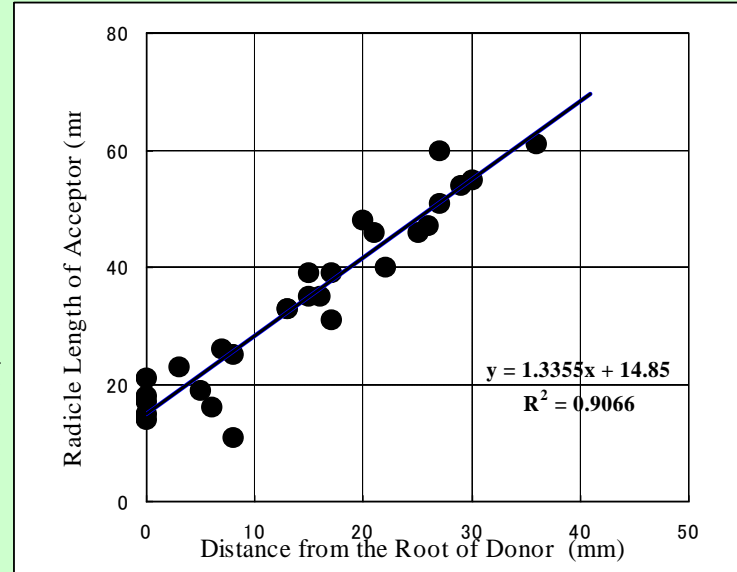
1 Plant Box Method

for root exudates (Mixed planting)



Root
zone
separating
tube

- Sand Culture for 1-2 month
- **Plant Box (for tissue culture)**
- Agar Medium (no nutrients)



Dr. Fujii, 1991

Fujii, Y. et al., Plant-Box Method: A Specific Bioassay to Evaluate Allelopathy through Root Exudates. Allelopathy, New Concepts and Methodology, Science Publisher, 39-56 (2007).

Table 1. Allelopathic activity Leguminosae Family by Plant Box Method

Scientific name

Pachyrhizus erosus
Mucuna pruriens var.
Mimosa invisa
Crotalaria zanzibarica
Medicago scutellata
Galega orientalis
Coronilla varia
Medicago arabica
Trifolium album
Medicago lupulina
Phaseolus vulgaris
Vicia villosa var. *dasy*
Abrus praecatorius
Canavalia ensiformis
Lupinus bicolor
Vicia villosa var. *villosa*
Melilotus albus
Dolicos lablab
Vicia sativa
Pueraria lobata
Crotalaria juncea
Trigonella foenum-gra
Vigna unguiculata
Cassia tora
Medicago sativa
Vicia faba
Onobrychis viciifolia
Pachyrhizus tuberosus
Phaseolus vulgaris
Vigna angularis
Trifolium incarnatum
Cajanus cajan
Crotalaria spectabilis
Pisum sativum
Cicer arietinum
Lathyrus odoratus
Amorpha fruticosa
Lathyrus sativus
Lupinus albus
Arachis hypogaea
Calopogonium mucun
Desmodium paniculat
Vigna unguiculata sub
Mimosa pudica
Vicia angustifolia var. *segetalis*

Allelopathic Leguminous Plants

Pachyrhizus

Mucuna

Mimosa

Crotalaria

Medicago

Vicia

Sensitive plant 55
 Karasunoendou 55



Mucuna



Mimosa

Plant Box Method

Table Allelopathic activity of Poaceae family evaluated by Plant Box Method (selected results)

Scientific name

Hordeum vulgare (Barley, cv. HVG-1)

Avena strigosa (Wild Oat)

Triticum polonicum (Polish Wheat)

Avena sativa (Oat, Hokuren)

Avena murphyi (Wild Oat)

Secale cereale (Rye, Samusa-shirazu)

Avena barbata (Wild Oat)

Triticum compactum (Club Wheat)

Avena longiglumis (Wild Oat)

Triticum spelta (Spelta Wheat)

Triticum sp. x Secale sp. (Triticale)

Avena orientalis (Wild Oat)

Chloris gayana (Rhodesgrass)

Avena wiestii (Wild Oat)

Avena byzantina (Byzantine Oat)

Hordeum vulgare (Barley, Sansyu)

Phalaris tuberosa (Hardinggrass)

Triticum aestivum (Common Wheat, Mulchmugi)

Anthoxanthum odoratum (Sweet Vernalgrass)

Festuca rubra (Chewing Fescue)

Panicum coloratum (Coloured Guinea Grass)

Avena fatua (Common wild oat)

Triticum aestivum (Common Wheat, cv. Norin 61)

Allelopathic Crops

Barley (*Hordeum vulgare*)

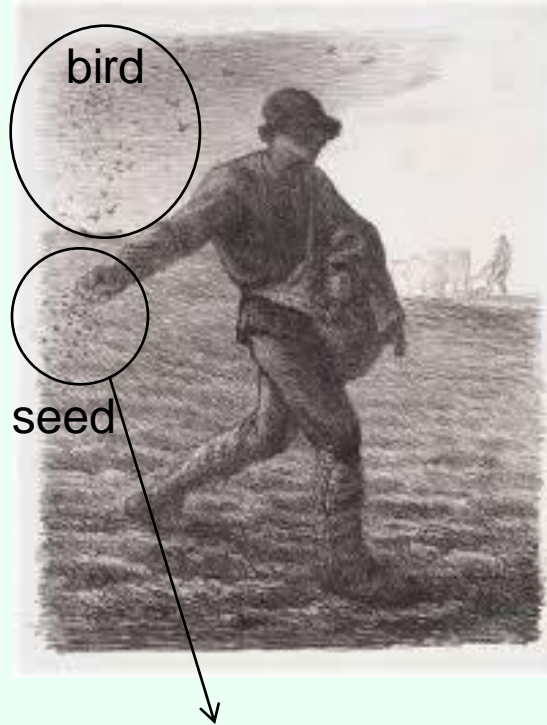
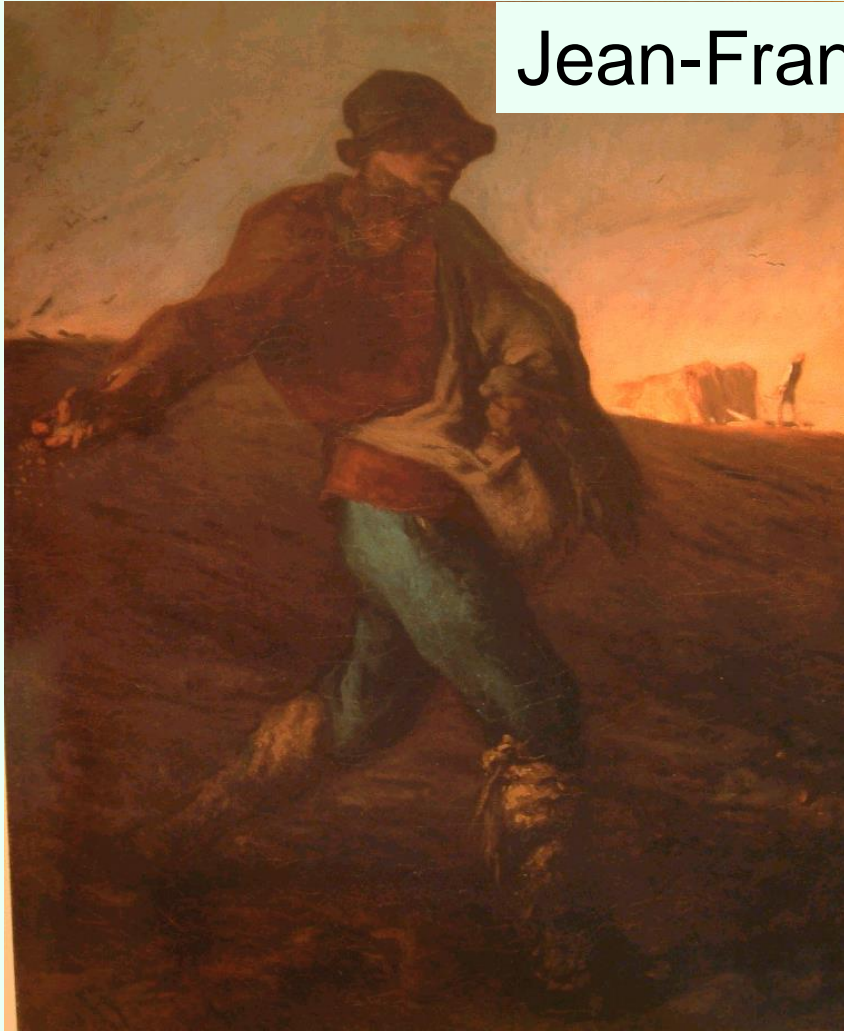
Oats (*Avena* sp.)

Rye (*Secale cereale*)

Wheat (*Triticum* sp.)

**These upland crops are
originally resistant for weed**

Jean-François Millet, Le semeur (1850)



We Japanese
love Millet !

Wheat wheat has allelopathic activity
amount of seeding rate = 3 to 5 times
For bird loss and weed suppression

Table Allelopathic activity of Triticum family
evaluated by Plant Box Method

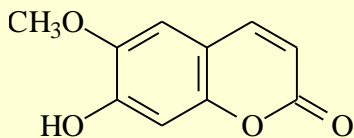
Scientific name	Radicle growth (%)
<i>Triticum polonicum</i> (Polish Wheat)	13.2
<i>Triticum compactum</i> (Club Wheat)	19.5
<i>Triticum spelta</i> (Spelta Wheat)	22.4
<i>Triticum sp. x Secale sp.</i> (Triticale)	22.6
<i>Triticum aestivum</i> (Common Wheat, cv. Mulchmugi)	28.0
<i>Triticum aestivum</i> (Common Wheat, cv. Norin 61)	33.5
<i>Triticum monococcum</i> (Einkorn Wheat)	34.3
<i>Triticum durum</i> (Durum Wheat)	37.3
<i>Triticum dicoccum</i> (Emmer Wheat)	40.5

“MACE” is King variety in AU now
(by Cristine Zaicou-Kunesch)
At today’s morning session

Proposal
for Project
Breeding of
Weed Resistant Wheat
with
high allelopathic
activity !

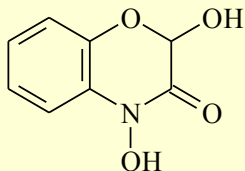
Allelochemicals reported from **Oats, Rye, Wheat, Barley**

Oat



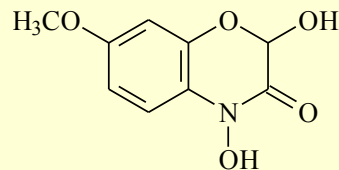
7-Hydroxy-5-methoxycoumarin
(**Scopoletin**)

Rye



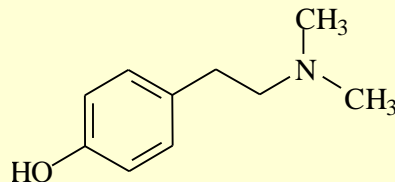
2,4-dihydroxy-
1,4-benzoxazine-3-one
(**DIBOA**)

Wheat



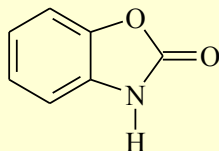
2,4-dihydroxy-7-methoxy-
1,4-benzoxazine-3-one
(**DIMBOA**)

Barley

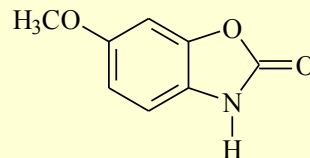


N,N-dimethyltyramine
(**Hordenine**)

Many other chemicals are reported, but I guess these traditional compounds are important as allelochemicals based on **TA**



2-benzoxazolinone
(**BOA**)

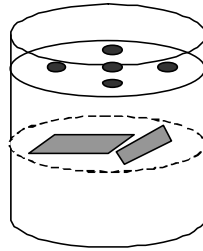
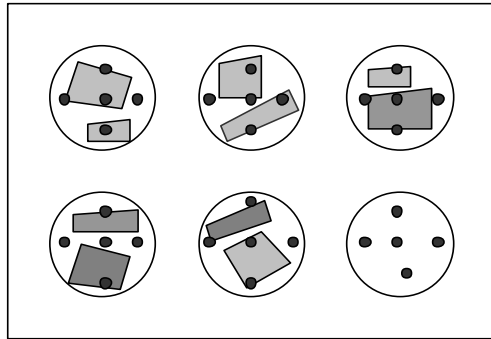


6-methoxy-
2-benzoxazolinone
(**MBOA**)

As for **“TA”**,
Total Activity,
Will explain later.

1. **Belz**, G.R. and **Hurle**, K., Dose-Response - A Challenge for Allelopathy? *Nonlinearity Biol. Toxicol. Med.* 3 (2), 173–211 (2005)
2. **Liu**, D. L. and **Lovett**, J. V., Biologically active secondary metabolites of barley. II. Phytotoxicity of barley allelochemicals, *J. Chem. Ecol.* 19, 2231-2244 (1993)

2. Sandwich Method *for leaf leachates (Mulching, Litter)*



6 well multi-dish
make sandwich by agar
→ 10 or 50 mg d.w./10 cm²

For allelopathy by fallen leaves and litters
fallen leaves are constant

(3 ton / ha / year) = 30 mg d.w./10 cm²

Dr. Fujii, 1991

Fujii, Y. et al., Assessment method for allelopathic effect from leaf litter leachates.
Weed Biology and Management, 4(1), 1923 (2004)

Table 3. Allelopathic activity by Sandwich Method

Scientific name	Radicle	Hypocotyl
<i>Cymbopogon citratus</i>	0**	0**
<i>Derris scandens</i>	0**	0**
<i>Piper betle</i>	3**	18**
<i>Tamarindus indica</i>	3**	26*
<i>Gliricidia sepium</i>	5**	19*
<i>Sesbania grandiflora</i>	11*	45*
<i>Acacia farnesiana</i>	15*	37
<i>Duranta repens</i>	17*	51
<i>Diospyros mollis</i>	23*	22
<i>Afgekia sericea</i>	38	120
<i>Ipomea pes-caprae</i>	39	90
<i>Jatropha integerrima</i>	44	122
<i>Melia azedarach</i>	52	115
<i>Citharexylum spinosum</i>	70	60
<i>Molineria latifolia</i>	75	167
<i>Passiflora coccinea</i>	86	117
<i>Calophyllum inophyllum</i>	94	146
<i>Amhersita nobilis</i>	96	127
<i>Cynometra cauliflora</i>	101	113
<i>Litchi chinensis</i>	113	115

Radicle and hypocotyl means % growth of lettuce seedling to the control (in agar medium). ** after the data means inhibitory activity stronger than standard deviation value of 60, and * means 55.

Sandwich Method



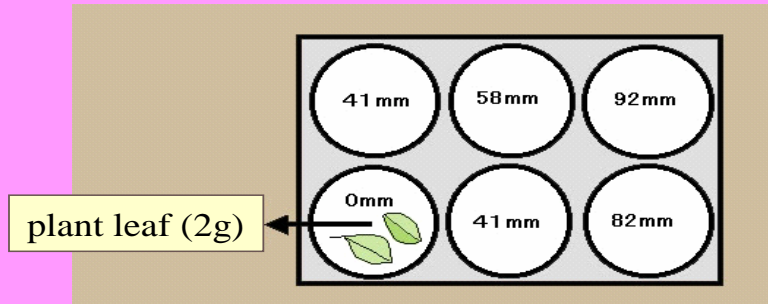
Cymbopogon



Tamarindus

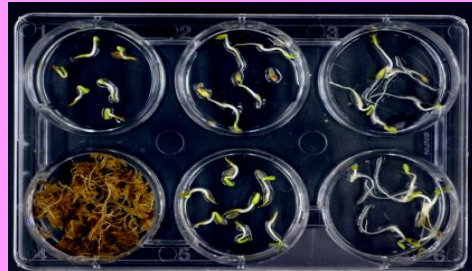
Fujii, Y. et al., Screening of 239 medicinal plant species for allelopathic activity using the sandwich method. *Weed Biology and Management*, 3(4), 233-241 (2003)

3. Dish Pack Method *for volatile allelochemicals*

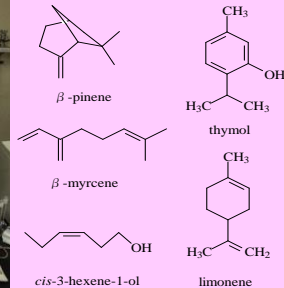


6 well multi-dish
sealed by tape

→ measure after 3 days



Volatile
chemicals
were
analyzed
by GC-MS

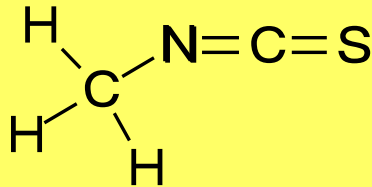


Fujii, Y. et al., Dish pack method: a new bioassay for volatile allelopathy. In Proceedings of the 4th World Congress on Allelopathy, August 21 -26, 2005, Wagga Wagga, Australia, 493-497 (2005).

Allelopathic plant for biofumigation



Cleome from Takii seed Co.



Methyl Isothiocyanate

Cleome

Cleome spinosa

Strongest activity by Dish Pack Method

Allelochemical : Methyl Isothiocyanate (MITC)

EC100 for lettuce is 18 ppm

Kill most species of nematode

Concentration of MITC in Cleome	
	ppm
Leaf (fresh)	39
Root (fresh)	15
Seed pod	16
Seed	14
Seed (macerated)	133

Table Database for Allelopathic activity evaluated by PB, SW, and DP method

<i>Abutilon theophrasti</i>	イデビ	アオイ科	31	105	21	1	<i>Echinochloa frumentacea</i>	イネ科	38	109	55	0	<i>Melilotus officinalis</i>	セイヨウシロツラギ	マメ科	2	62	19	3
<i>Aegilops cylindrica</i>	ヤギムギ	イネ科	60	111	27	1	<i>Echinochloa hispidula</i>	イネ科	46	109	68	0	<i>Melilotus sulcata</i>		マメ科	1	37	30	2
<i>Agropyron repens</i>		イネ科	44	119	60	0	<i>Echinochloa oryzoides</i>	ノゲノタイシビエ	イネ科	25	94	30	1	<i>Mentha arvensis</i>	シソ科	32	75	97	1
<i>Agrostis canina</i>	ヒメスカタネ	イネ科	36	97	29	0	<i>Echinochloa utilis</i>		イネ科	22	131	54	1	<i>Myosotis arvensis</i>	ムラサキ科	64	88	37	0
<i>Agrostis capillaris</i>	イネコノハダサ	イネ科	42	90	24	1	<i>Echium italicum</i>	ムラサキ科	73	89	41	0	<i>Nicotiana sylvestris</i>	ナス科	71	100	40	0	
<i>Agrostis castellana</i>		イネ科	46	106	43	0	<i>Echium plantagineum</i>	シヤゼンムラサキ	28	129	43	0	<i>Oenothera crocata</i>	セリ科	25	95	93	1	
<i>Agrostis olivacea</i>		イネ科	43	88	43	0	<i>Echium vulgare</i>	シヤゼンムラサキ	30	107	46	0	<i>Oenothera erythrosealae</i>	アホバノ科	65	113	47	0	

Evaluation of Allelopathy by specific bioassays
and evaluated about 4,000 species in 20 years



- 1) Medicinal Plants
 - 2) Tropical Plants
 - 3) Endangered Plants
 - 4) Invasive Alien Plants
 - 5) Oats, Barley, Wheat
- } are Allelopathic

(My dream) Database for Allelopathic activity evaluated by PB, SW, DP method

2) Isolation of Allelochemicals based on Total Activity

Chromatography (TLC, HPLC ...)

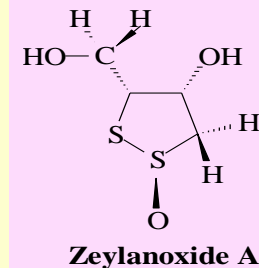
試料添加

薄層クロマトグラフィー用
シリカゲルプレート

溶媒(油)で展開

さらに展開

さらに展開
良好な分離！



Structure



Solvent Extraction



GC-MS, MS,
NMR, IR, ESR,
X-ray analysis,
etc

Conventional Method for Isolation of natural chemicals

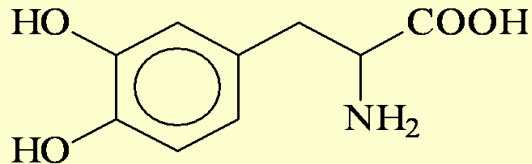
New strategies for isolation of allelochemicals in agriculture

Comparison of **Total Activity** and **Specific Activity**

Target	Compounds	Index (g-weight base)
Direct use of Allelopathy	Allelochemicals in action <i>in situ</i>	<div>Total Activity = $1/EC_{50} \times \text{Concentration in Plants}$ $1/(\text{kg/L}) \times (\text{kg/L in fresh weight})$ $= (\text{L/kg}) \times (\text{kg/L}) = (\text{no unit})$ high value ~ high activity</div>
Use of Natural Chemicals	Bioactive Chemicals with high specific activity	<div>Specific Activity = EC_{50} (50% inhibitory Concentration) $(\text{kg/L}) \sim \text{ppm}$ low ppm ~ high activity</div>

Isolation of L-DOPA from Velvet bean

Velvet bean (*Mucuna pruriens*)



L-DOPA (L-3,4-Dihydroxyphenylalanine)

High concentration
10g/kg in fresh leaves, root
50-100g/kg in seeds

Total activity
= 200

Total activity = 200

**Endemic plant
to Himalayas
Distributed in
Sino-Japan Area**

**Used as food
in Japan from
more than
1000 years ago
(but now
disappearing)**

Fujii Y, Shibuya T. and Yasuda T. L-3,4-dihydroxyphenylalanine as an allelochemical candidate from *Mucuna pruriens* (L.) DC. var. *utilis*. *Agricultural and Biological Chemistry* 55, 617-618. (1991)

Isolation of cyanamide from hairy vetch

Hairy vetch (*Vicia villosa*)



Cyanamide was already known as synthetic fertilizer, but never isolated as natural chemical

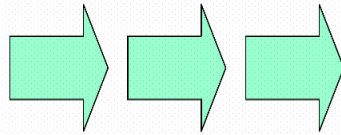
Endemic plant to central Asia,

now we are recommending to Japanese farmers for weed control



Cyanamide

- Antibacterial activity
- fungicidal activity
- herbicidal activity



Biochemical mechanisms



Ammonia

- Nitrogen fertilizer

Total activity
= 35 - 100

Kamo T, Hiradate S. and Fujii Y. First isolation of natural cyanamide as a possible allelochemical from hairy vetch *Vicia villosa*., *J. Chem. Ecol.*, 29 (2), 275-284 (2003)

3) Practical use of Allelopathic Cover Crops

Mucuna (velvetbean) (*Mucuna pruriens*)

for Green manure, **Food, Medicinal plants**, and Cover Crop

Fujii, Y. Allelopathy of Velvetbean: Determination and Identification of Allelopathic Substances. H. G. Cutler and S. Cutler (Eds), Biologically Active Natural Products: Agrochemicals, CRC Press, pp.33-47 (1999)

Velvetbean : *Mucuna pruriens* var. *utilis*

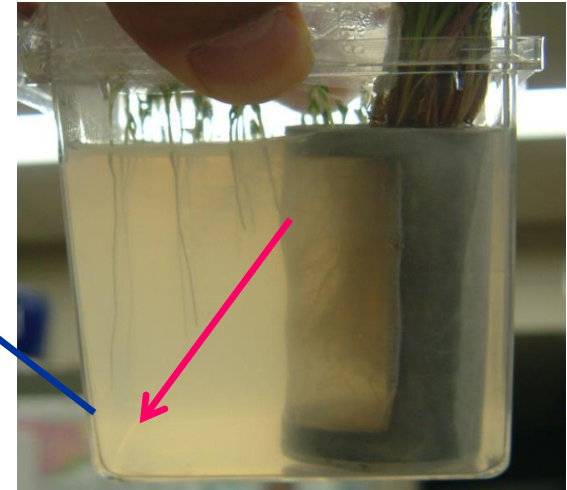


Legume, rich in protein

Table 1. Assessment of Allelopathic Activity of Leguminous Cover Plants by Plant Box Method

Scientific name (English name)	%*1	Criteria*2
Mucuna pruriens var. utilis (Mucuna)	87	★★★
Vicia faba (Broad Bean)	81	★★
Vicia villosa (Hairy Vetch)	80	★★
Calopogonium mucunoides	78	★★
Melilotus albus (White Sweet Clover)	77	★★
Vicia sativa (Common Vetch)	75	★★
Medicago rugosa	74	★★
Canavalia ensiformis (Jack Bean)	72	★★
Pueraria phaseoloides (Tropical Kudzu)	71	★★
Vigna angularis (Azuki bean)	69	★
Medicago sativa (Alfalfa)	68	★
Trifolium incarnatum (Crimson Clover)	64	★
Tephrosia candida (White Tephrosia)	63	★
Cajanus cajan (Pigeon Pea)	62	★
Latyrus sativus (Grass Pea)	59	★
Cicer arietinum (Chickpea)	56	★
Vigna radiata (Mung Bean)	53	
Stylosanthes hamata (Stylo)	52	
Arachis hypogaea (Peanut)	51	
Trifolium pratense (Red Clover)	47	
Crotalaria juncea (Sunn Hemp)	43	
Astragalus sinicus (Chinese Milk Vetch)	41	
Lupinus albus (White Lupine)	40	
Trifolium subterraneum (Sub Clover)	30	
Trifolium repens (White Clover)	28	
Glycine max (Soybean)	24	

Plant Box Method (for root exudates)



- *1 Inhibition (%) means allelopathic activity measured by the Plant Box Test. All data are compared to the control and 100 means complete inhibition.
- *2 Criteria for the allelopathic activity are ;
 ★★★ ; >85 % , ★★ ; 70 to 84% ,
 ★ : <69 % , respectively.

Table Weed population in continuous cropping fields

Crop	Treatment	Weed population (g dry weight per m ²)	Weed species observed ⁶⁾
Upland Rice	3yr.c ¹⁾	5.11(49.4) ⁴⁾	1), 3), 5), 6), 7), 8), 9), 10), 11) ⁵⁾
Egg plant	3yr.c	16.82(40.1)	1), 2), 3), 5), 6), 7), 8), 9), 10), 11), 12), 13), 14)
Tomato	3yr.c	4.92(64.9)	1), 5), 6), 9), 12), 13), 17)
Velvetbean	2yr.c	0.00(0.0)	no emergence
Fallow	3yr.f ³⁾	0.97(37.3)	1), 2), 6), 10), 12), 13), 15), 16)

1) Continuous cropping for 3 years.

2) Cultivated for 1 year, followed by fallow next year(test year).

3) Fallow for 3 years, without fertilizer.

4) Numbers in parenthesis are percentages of chickweed, a dominant species.

5) Species appeared in each plot: 1) Sticky chickweed (*Cerastium glomeratum*),

2) 'Mimnagusa' (*Cerastium vulgatum* var. *augustifolium*), 3) Annual fleabane

(*Erigeron annuus*), 4) Philadelphia fleabane (*Erigeron philadelphicus*),

5) Starwort (*Stellaria alsine* var. *undulata*), 6) Floating foxtail (*Alopecurus*

geniculatus), 7) Narrowleaf vetch (*Vicia angustifolia*), 8) Flexuosa bittercress

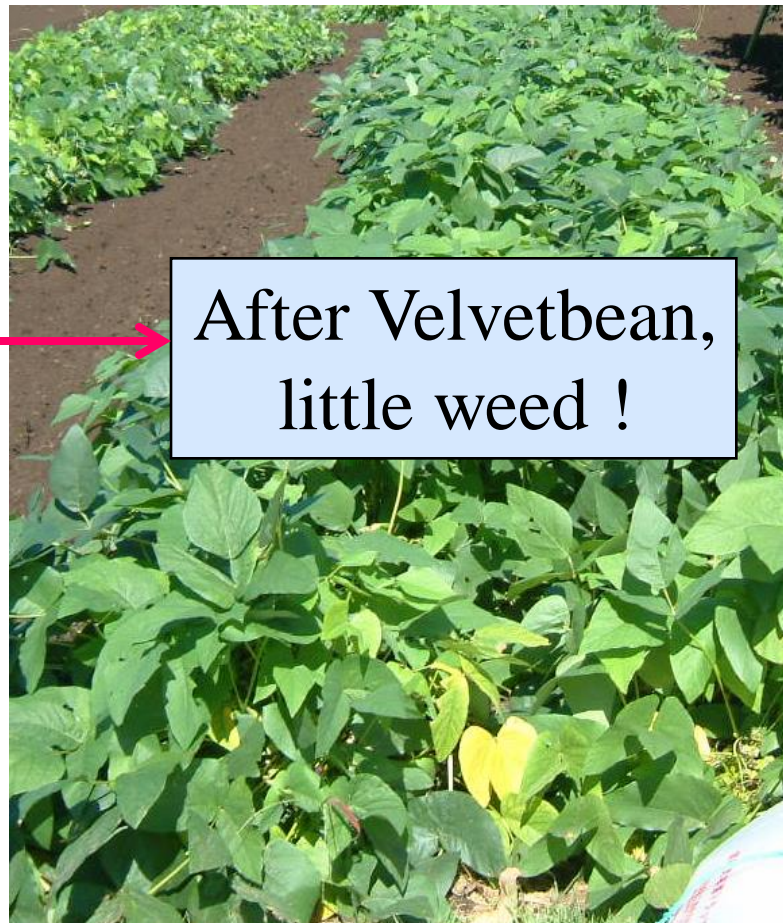
(*Cardamine flexuosa*), 9) 'Inugarashi' (*Rorippa atrovirens*), 10) Common dandelion

(*Taraxacum officinale*), 11) Japanese mugwort (*Artemisia princeps*), 12) Canadian

fleabane (*Erigeron canadensis*), 13) 'Hahakogusa' (*Gnaphalium affine*), 14) Blady

grass (*Imperata cylindrica*), 15) Meadowgrass (*Poa annua*), 16) Creeping wood-

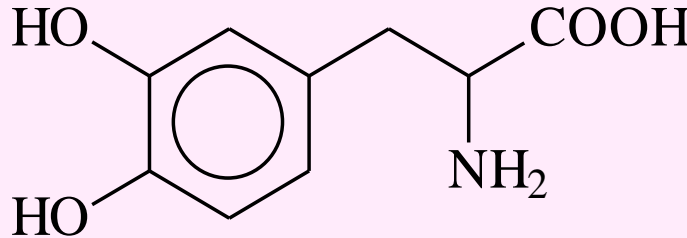
sorrel (*Oxalis corniculata*), 17) Shepherd's-purse (*Capsella bursa-pastoris*),



Fujii, Y. Shibuya, T. and Usami, Y. Allelopathic effect of *Mucuna pruriens* on the appearance of weeds, *Weed Res, Japan* 36, 43-49 (1991)

Allelochemical in *Mucuna* (Velvetbean)

L-3,4-Dihydroxyphenylalanine (L-DOPA)



- ♦ Extremely high concentration in *Mucuna pruriens* **leaves, roots (0.5 to 1.5%) and seeds (5 to 8%) in fresh wt.**
- ♦ Relatively strong inhibition to broadleaf weeds ($EC_{50}=5-50\text{ppm}$), but not so active to grass (monocot.)

Fujii, Y., T. Shibuya, and T. Yasuda, L-3,4-Dihydroxyphenylalanine as an allelochemical candidate from *Mucuna pruriens*(L.) DC.var.*utilis*. Agric.Biol.Chem. 55(2), 617-618 (1991)

How to use *Mucuna*

- 1) Green Manure
- 2) Vegetable (Young pod)
- 3) Seeds as food, but after proper Cooking.
- 4) Medicine, as a source of L-DOPA



Hairy vetch (*Vicia villosa*)



for Green manure and Weed control

Fujii, : Screening and Future Exploitation of Allelopathic Plants as Alternative Herbicides with Special Reference to Hairy Vetch. R. K. Kohli, H. P. Singh, and D. R. Batish (Eds), Allelopathy in Agroecosystems, pp.257-275 (2001)

First observation of strong weed suppression on the field, August, 1990



Table Effect of Cover Crops on Weed Control in Abandoned Paddy Field

; First Trial on the Experimental Station (1992-1993)

Cover Crop (English Name)	W (%)* ¹	Crop Yield [g m ⁻²]
First Sampling (May 7, 1993)		
Control(No-weeding)	0 a ^{*2}	----
<i>Astragalus sinicus</i> (Chinese Milk Vetch)	82 b	431 a
<i>Vicia villosa</i> (Hairy Vetch)	99 b	584 ab
<i>A. sativa</i> + <i>V. villosa</i> (Oat and Hairy Vetch Mix.)	99 b	730 b
Second Sampling (June 10, 1993)		
Control (No-weeding)	0 a	----
<i>Astragalus sinicus</i> (Chinese Milk Vetch)	59 b	135 a
<i>Vicia villosa</i> (Hairy Vetch)	100 c	147 b
<i>A. sativa</i> + <i>V. villosa</i> (Oat and Hairy Vetch mix.)	100 c	137 a

*1 Percentage of weed control. 100 % means complete control. Dry weight of weeds in the control plot are 281 (First) and 155 (Second) g m⁻².

Fujii, Y. Screening and future exploitation of allelopathic plants as alternative herbicides with special reference to hairy vetch, *J. Crop Prod.*, 4 (2) , 257-275 (2001)

32

Orange Farm

Mr.Ochi
(Ehime Pref.)

1991年



Local Government recommend (Minami-Ashigara city)



Hairy vetch is now planted 10,000 ha in Japan
(No 2 as cover crop, next to traditional Milk vetch)

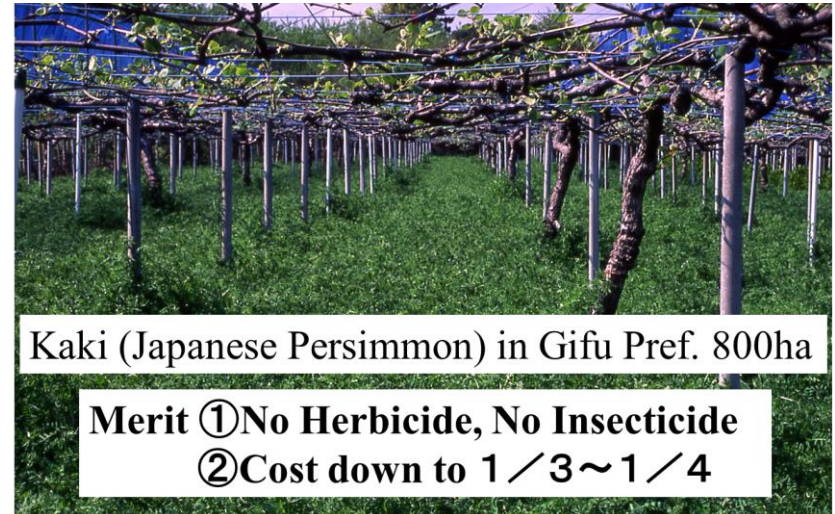
No herbicide, no fertilizer, only Vetch (Kiwi Fruits)



Mr. Ishiwata
Odawara City

My Recommendation at 1991 →
Starting in his Fruit garden
after 10 years

No herbicide, No fertilizer, No
organic materials than vetch →
now spreading all around Japan in
Orchard

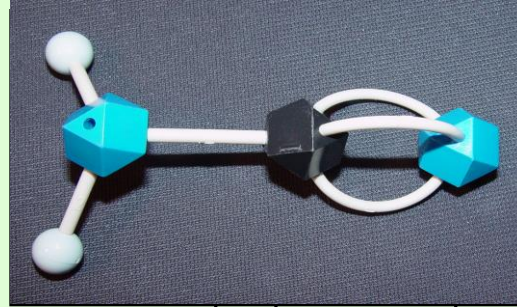


Kaki (Japanese Persimmon) in Gifu Pref. 800ha

Merit ①No Herbicide, No Insecticide
②Cost down to 1/3~1/4

Identification of Cyanamide as Allelochemical from Hairy vetch

★ Cyanamide



★ known as active constituents of synthetic fertilizer (Calcium cyanamide) **First finding from plants**

Kamo, T., Hiradate, S. and Fujii, Y.: First isolation of natural cyanamide as a possible allelochemical from hairy vetch *Vicia villosa* . *J. Chemical Ecol.* 29 (2), 275-283 (2003)

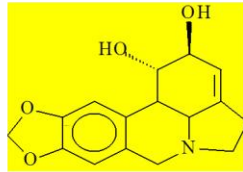


Hairy Vetch used as
cover crop in Japan
more than 10,000 ha
(estimated from 300 tons seed)
No.2 as cover crop
...after 18 years

Other Promising Allelopathic Cover Plants

Red Spider Lily *Lycoris radiate*

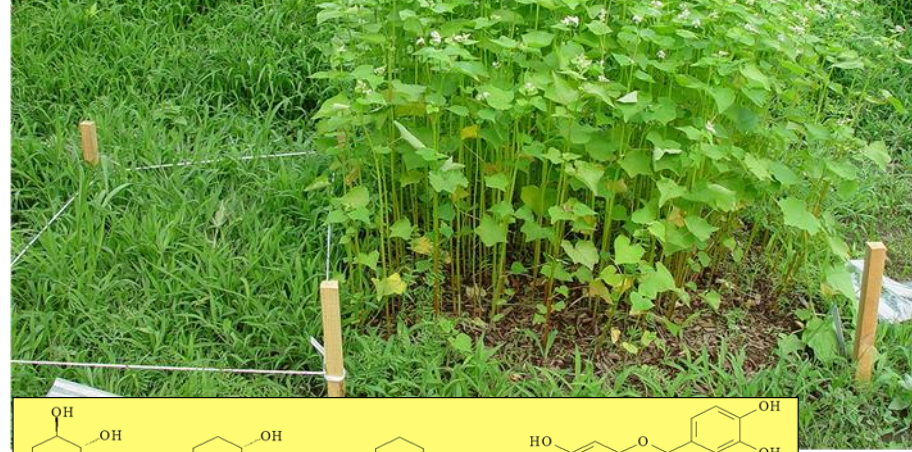
- Japanese traditional plants originally came from **South China (Yunnan)**
- Farmers notice allelopathy



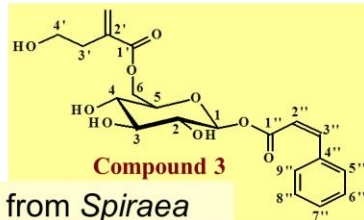
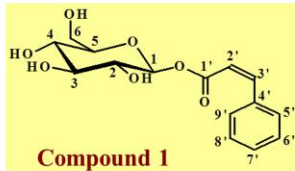
Allelochemical: **Lycorine**

Iqbal, Z., Nasir, H., Hiradate, S. and Fujii, Y. Plant growth inhibitory activity of *Lycoris radiate* Herb. and possible involvement of lycorine as an allelochemical. *Weed Biology and Management*, 6 (4), 221-227 (2006).

Buckwheat *Fagopyrum esculentum*

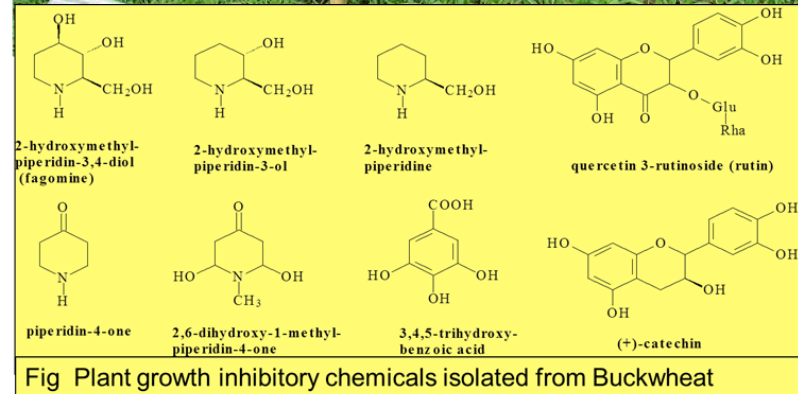


Thunberg spiraea Yuki-yanagi 雪柳 (*Spiraea thunbergii*)



Isolation of *cis*-cinnamic acid from *Spiraea*

Hiradate, S., S. Morita, H. Sugie, Y. Fujii, and J. Harada, Phytotoxic *cis*-cinnamoyl glucosides from *Spiraea thunbergii*. *Phytochemistry*, 65(6), 731-739 (2004)



Iqbal, Z., Hiradate, S., Noda, A., Isojima, S. and Fujii, Y. : Allelopathy of buckwheat: Assessment of allelopathic potential of extract of aerial parts of buckwheat and identification of fagomine and other related alkaloids as allelochemicals. *Weed Biology and Management* 2, 110-115 (2002)

Ground Cover Plants

Resistant for insects,
disease & weeds

Perfume, Medicinal chemicals

Beautiful flowers

with Allelopathic activity

Honey

Useful
as
Forage

O₂ ↑

Suppress weeds

N₂ fixation

Prevent
Erosion

Food (Fruit,
Seed, Tuber...)

Cultivate
soil by root
system

Keep soil
moisture

Green manure by
Nitrogen fixation and
organic carbon

Clean polluted soil by
toxic chemicals

Resistant for nematoda,
and soil disease

Phyto-Lark : Ground Cover plants with allelopathy

Three key messages

- **Specific bioassays for Allelopathy**, and evaluated about 4,000 plants.
- New Allelochemicals Isolated by new methodology, “**Total Activity Method**”
- **Ground Cover Plants with allelopathy**
(= Phytolark) provides natural and environmentally friendly way of managing weeds without herbicides.



Questions?

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